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**ASSISTANT COMMISSIONER FOR PATENTS  
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Washington, D.C. 20231**

Attorney Docket No. 18062L-000110

"Express Mail" Label No. EL525626338US

Date of Deposit: August 28, 2000

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By:

Stuart McLeish

Submitted herewith for filing under 37 CFR 1.53(b) is the  
☐ patent application of  
☒ continuation patent application of  
☐ divisional patent application of  
☐ continuation-in-part patent application of

Inventor(s)/Applicant Identifier: PETER C. SIMPSON et al.

## For: MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS DEVICE AND METHOD

☒ This application claims priority from each of the following Application Nos./filing dates:

U.S. application no. 08/965,738 filing date November 7, 1997

the disclosure(s) of which is (are) incorporated by reference.

## Enclosed are:

☒ 28 total page(s) of specification, claims & abstract☒ 7 page(s) of claims☒ 1 page of Abstract☒ 11 sheet(s) of ☒ formal ☐ informal drawing(s).☒ A copy of the signed assignment from the prior application of the invention to The Regents of the University of California☒ A copy of the signed Declaration from the prior application.☒ A copy of the signed Power of Attorney by Assignee from the prior application.☒ A copy of the signed verified statement to establish small entity status under 37 CFR 1.9 and 37 CFR 1.27 was filed in the prior application and small entity status is still proper and desired (copy enclosed).☒ A copy of the Revocation Substitute Power of Attorney filed in the prior application.☒ Preliminary Amendment.☒ Return Postcard

	(Col. 1)	(Col. 2)	
FOR:	NO. FILED	NO. EXTRA	
BASIC FEE			
TOTAL CLAIMS	14 - 20	= *0	
INDEP. CLAIMS	6 - 3	= *3	
<input type="checkbox"/> MULTIPLE DEPENDENT CLAIM PRESENTED			

\* If the difference in Col. 1 is less than 0, enter "0" in Col. 2.

## SMALL ENTITY

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x \$9.00 =	\$0.00
x \$39.00 =	\$117.00
+ \$130.00 =	
TOTAL	\$462.00

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OR	\$690.00
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OR	x \$78.00 =
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Respectfully submitted,  
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David Heckadon

(Granted Limited Recognition under 37 CFR §10.9(b)  
(see enclosed Limited Recognition Document)  
Attorneys for Applicant

Applicant or Patentee: Peter C. Simpson, et al.

Serial or Patent No.: 08/965,738

Filed or Issued: November 7, 1997

For: MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS DEVICE AND METHOD

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS  
(37 CFR 1.9(f) and 1.27(d)) - NONPROFIT ORGANIZATION

I hereby declare that I am an official empowered to act on behalf of the nonprofit organization identified below:

Name of Organization: The Regents of the University of California  
Address of Organization: 300 Lakeside Drive, 22nd Floor, Oakland, California 94612-3550  
Type of Organization:

- ☒ UNIVERSITY OR OTHER INSTITUTION OF HIGHER EDUCATION  
☐ TAX EXEMPT UNDER INTERNAL REVENUE SERVICE CODE (26 USC 501(a) and 501(c)(3))  
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(NAME OF STATE: California )  
(CITATION OF STATUTE: )  
☐ WOULD QUALIFY AS TAX EXEMPT UNDER INTERNAL REVENUE SERVICE CODE (26 USC 501(a) and 501(c)(3)) IF  
LOCATED IN THE UNITED STATES OF AMERICA  
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AMERICA IF LOCATED IN THE UNITED STATES OF AMERICA  
(NAME OF STATE: California )  
(CITATION OF STATUTE: )

I hereby declare that the nonprofit organization identified above qualifies as a nonprofit organization as defined in 37 CFR 1.27(e) for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code with regard to the invention entitled MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS DEVICE AND METHOD by inventor(s) Peter C. Simpson, Richard A. Mathies, and Adam T. Woolley described in

- ☐ the specification filed herewith.  
☒ application serial no. 08/965,738, filed November 7, 1997  
☐ patent no. \_\_\_\_\_, issued \_\_\_\_\_.

I hereby declare that rights under contract or law have been conveyed to and remain with the nonprofit organization with regard to the above identified invention.

If the rights held by the nonprofit organization are not exclusive, each individual, concern or organization having rights to the invention is listed below\* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.90 or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

\*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

Full Name: The Regents of the University of California  
Address: 300 Lakeside Drive, 22nd Floor, Oakland, California 94612-3550

☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☒ NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

Name: William A. Hoskins

Title: Director, University of California at Berkeley

Address: Office of Technology Licensing, 2150 Shattuck Avenue, Suite 510  
Berkeley, California 94720

Signature: [Signature]

Date: May 4, 1998

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Washington, D.C. 20231

By: 

Stuart McLeish

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

In re application of:

PETER C. SIMPSON et al.

Application No.: Unassigned

Filed: Herewith

For: MICROFABRICATED  
CAPILLARY ARRAY  
ELECTROPHORESIS DEVICE  
AND METHOD

Examiner: Unassigned

Art Unit: Unassigned

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents  
Washington, D.C. 20231

Sir:

Prior to examination of the above-referenced application, please enter the following amendments and remarks.

IN THE SPECIFICATION:

After the title, please insert the following paragraph:

--This application is a continuation of, and claims the benefit of priority from U.S. application No. US 08/965,738, filed on November 7, 1997, the full disclosure of which is incorporated herein by reference.--

IN THE CLAIMS:

Please cancel claims 1-36. Please substitute new claims 1-14.

--1. A capillary array electrophoresis plate, comprising:

2 a separation channel having a cathode reservoir at one and an anode  
3 reservoir at an opposite end; and  
4 an injection channel having a first leg and a second leg, the first leg  
5 connected at one end to a plurality of sample reservoirs and at the other end to the separation  
6 channel, and the second leg connected at one end to the separation channel and at the other end  
7 to a waste reservoir.

1 2. The capillary array electrophoresis plate of claim 1, wherein the first and  
2 second legs of the injection channel are disposed collinear with one another.

1 3. The capillary array electrophoresis plate of claim 1, wherein the first leg  
2 of the injection channel is connected at one end to a loading channel connected to the plurality  
3 of sample reservoirs.

1 4. A capillary array electrophoresis plate, comprising:  
2 an array of separation channels, each separation channel having a  
3 cathode reservoir at one end and an anode reservoir at an opposite end; and  
4 an array of injection channels, each injection channel having a first leg  
5 and a second leg, the first leg connected at one end to a plurality of sample reservoirs and at the  
6 other end to one of the separation channels, and the second leg connected at one end to one of  
7 the separation channels and at the other end to a waste reservoir.

1 5. The capillary array electrophoresis plate of claim 4, wherein  
2 the cathode reservoirs are multiplexed.

1 6. The capillary array electrophoresis plate of claim 4, wherein  
2 the anode reservoirs are multiplexed.

1 7. The capillary array electrophoresis plate of claim 4, wherein  
2 the waste reservoirs are multiplexed.

1 8. A method of sequentially loading a plurality of different samples onto an  
2 electrophoretic separation channel, comprising:

3 providing a capillary array electrophoresis plate, comprising:  
4 a separation channel having a cathode reservoir at one and an  
5 anode reservoir at an opposite end; and  
6 an injection channel having a first leg and a second leg, the first  
7 leg connected at one end to a plurality of sample reservoirs and at the other end to the  
8 separation channel, and the second leg connected at one end to the separation channel and at  
9 the other end to a waste reservoir;  
10 moving a first sample from a first sample reservoir through first leg of  
11 the injection channel and into the separation channel; and subsequently,  
12 electrophoretically separating the first sample in the separation channel;  
13 and subsequently,  
14 moving a second sample from a second sample reservoir through first  
15 leg of the injection channel and into the separation channel; and subsequently,  
16 electrophoretically separating the second sample in the separation  
17 channel.

1 9. A method of sequentially loading a plurality of different samples onto an  
2 electrophoretic separation channel, comprising:

3 providing a capillary array electrophoresis plate, comprising:  
4 an array of separation channels, each separation channel having a  
5 cathode reservoir at one end and an anode reservoir at an opposite end; and  
6 an array of injection channels, each injection channel having a  
7 first leg and a second leg, the first leg connected at one end to a plurality of sample reservoirs  
8 and at the other end to one of the separation channels, the second legs connected at one end to  
9 one of the separation channels and at the other end to a waste reservoir;  
10 moving a plurality of first samples from the plurality of first sample  
11 reservoirs through the plurality of first legs of the injection channels and into the plurality of  
12 separation channels; and subsequently,  
13 electrophoretically separating the plurality of first samples in the  
14 separation channel; and subsequently,

moving a plurality of second samples from the plurality of second sample reservoirs through the plurality of first legs of the injection channels and into the plurality of separation channels; and subsequently, electrophoretically separating the plurality of second samples in the separation channel.

10. A capillary array electrophoresis plate, comprising:  
a separation channel having a cathode reservoir at one and an anode reservoir at an opposite end; and  
an injection channel having a first leg and a second leg, wherein,  
the first leg is connected at one end to a first waste reservoir and at the other end to the separation channel, and a first plurality of sample reservoirs are connected to the first leg along the length of the first leg; and  
the second leg is connected at one end to a second waste reservoir and at the other end to the separation channel, and a second plurality of sample reservoirs are connected to the second leg along the length of the second leg.

11. A method of sequentially loading four different samples onto an electrophoretic separation channel, comprising:  
providing a capillary array electrophoresis plate, comprising:  
a separation channel having a cathode reservoir at one and an anode reservoir at an opposite end; and  
an injection channel having a first leg and a second leg, wherein,  
the first leg is connected at one end to a first waste reservoir and at the other end to the separation channel and a plurality of sample reservoirs are connected to the first leg along the length of the first leg; and  
the second leg is connected at one end to a second waste reservoir and at the other end to the separation channel and a plurality of sample reservoirs are connected to the second leg along the length of the second leg;  
moving a first sample from a first sample reservoir through first leg of the injection channel and into the separation channel; and subsequently,

15 electrophoretically separating the first sample in the separation channel.

1 12. The method of claim 11, further comprising:  
2 moving a second sample from a second sample reservoir through first  
3 leg of the injection channel and into the separation channel; and subsequently,  
4 electrophoretically separating the second sample in the separation  
5 channel; and subsequently.

1 13. The method of claim 11, further comprising:  
2 moving a third sample from a third sample reservoir through second leg  
3 of the injection channel and into the separation channel; and subsequently,  
4 electrophoretically separating the third sample in the separation channel;  
5 and subsequently.

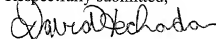
1 14. The method of claim 11, further comprising:  
2 moving a fourth sample from a second sample reservoir through second  
3 leg of the injection channel and into the separation channel; and subsequently,  
4 electrophoretically separating the fourth sample in the separation  
5 channel.--

CONCLUSION

In view of the foregoing, Applicants believe all claims now pending in this Application are in condition for allowance. The issuance of a formal Notice of Allowance at an early date is respectfully requested.

If the Examiner believes a telephone conference would expedite prosecution of this application, please telephone the undersigned at 650-326-2400.

Respectfully submitted,



David Heckadon

(Granted Limited Recognition under 37 CFR §10.9(b) -  
see enclosed Limited Recognition Document)

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By

Kimberly Badier  
Kimberly Badier

PATENT  
Attorney Docket No.: 18062L-000100  
UC Ref. No.: B98-020

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:

PETER S. SIMPSON et al.

Serial No.: 08/965,738

Filed: November 7, 1997

For: MICROFABRICATED  
CAPILLARY ELECTROPHORESIS  
DEVICE AND METHOD

Examiner: BEISNER, W.

Art Unit: 1744

Batch No.: L75

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Sir:

Enclosed please find a REVOCATION & SUBSTITUTION OF ATTORNEY for the  
above-identified patent application.

Respectfully submitted,

David R. Heckadon

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On JUNE 2000

TOWNSEND and TOWNSEND and CREW LLP

By:

NANCY A. PIZZO

**PATENT**

Attorney Docket No.: 18062L-000100

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

In re application of:

PETER C. SIMPSON et al.

Application No.: 08/965,738

Filed: November 7, 1997

For: MICROFABRICATED CAPILLARY ARRAY  
ELECTROPHORESIS DEVICE & METHOD

Examiner: BEISNER, W.

Art Unit: 1744

**REVOCATION & SUBSTITUTION  
OF ATTORNEY**

Assistant Commissioner for Patents  
Washington, D.C. 20231

Sir:

Pursuant to 37 C.F.R. § 1.36, UCB Office of Technology Licensing revokes all previous powers of attorney and hereby appoints the following to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

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**University of California, Berkeley**

Date: MAY 30 2000

William A. Hoskins  
Name: William A. Hoskins

Title: Director, UCB Office of Technology Licensing

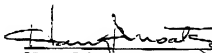
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**LIMITED RECOGNITION UNDER 37 CFR §10.9(b)**

David R. Heckadon is hereby given limited recognition under 37 CFR §10.9(b), as an employee of the Townsend and Townsend and Crew, law firm, to prepare and prosecute patent applications and to represent patent applicants wherein the patent applicants are clients of the Townsend and Townsend and Crew law firm, and wherein a registered practitioner who is a member of the Townsend and Townsend and Crew law firm is the attorney or agent of record. This limited recognition shall expire on the date appearing below, or when whichever of the following events first occurs prior to the date appearing below: (i) David R. Heckadon ceases to lawfully reside in the United States; (ii) David R. Heckadon's employment with the Townsend and Townsend and Crew law firm ceases or is terminated; or (iii) David R. Heckadon ceases to remain or reside in the United States on an H-1B visa.

This document constitutes proof of such recognition. The original of this document is on file in the Office of Enrollment and Discipline of the U.S. Patent and Trademark Office.

**Expires: November 23, 2000**

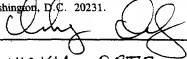
  
Harry I. Mdatz, Acting Director  
Office of Enrollment and Discipline

APPLICATION  
FOR  
UNITED STATES LETTERS PATENT

TITLE: MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS  
DEVICE AND METHOD

APPLICANT: PETER C. SIMPSON; RICHARD A. MATHIES; ADAM T.  
WOLLEY

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\_\_\_\_\_  
VICKY ORTIZ

MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS DEVICE AND METHOD

Statement as to Federally Sponsored Research

5 This invention was made with Government support under Grant No. DE-FG-91ER61125, awarded by the U.S. Department of Energy, and Grant No. HG01399, awarded by the National Institutes of Health. The Government has certain rights in the invention.

Background of the Invention

10 This invention relates to electrophoresis generally, and more particularly, to an apparatus and method for performing capillary array electrophoresis on microfabricated structures.

15 In many diagnostic and gene identification procedures such as gene mapping, gene sequencing and disease diagnosis, deoxyribonucleic acid (DNA), ribonucleic acid (RNA) or proteins are separated according to their physical and chemical properties. In addition to DNA, RNA or proteins, other small molecule analytes may also need to be separated.

20 One electrochemical separation process is known as electrophoresis. In this process, molecules are transported in a capillary or a channel which is connected to a buffer-filled reservoir. An electric field in the range of kilovolts is applied across both ends of the channel to cause the molecules to migrate. Samples are typically introduced at a high potential  
25 end and, under the influence of the electric field, move toward a low potential end of the channel. After migrating through the

channel, the separated samples are detected by a suitable detector.

Typically, electrophoretic separation of nucleic acids and proteins is carried out in a gel separation medium. Although slab gels have played a major role in electrophoresis, difficulties exist in preparing uniform gels over a large area, in maintaining reproducibility of the different gels, in loading sample wells, in uniformly cooling the gels, in using large amounts of media, buffers, and samples, and in requiring long run times for extended reading of nucleotides. Moreover, slab gels are not readily amenable to a high degree of multiplexing and automation. Recently, micro-fabricated capillary electrophoresis (CE) devices have been used to separate fluorescent dyes and fluorescently labeled amino acids. Additionally, DNA restriction fragments, polymerase chain reaction (PCR) products, short oligonucleotides and even DNA sequencing fragments have been effectively separated with CE devices. Also, integrated micro-devices have been developed that can perform polymerase chain reaction amplification immediately followed by amplicon sizing, DNA restriction/digestion and subsequent size-based separation, and cells sorting and membrane lysis of selected cells. However, these micro-fabricated devices only perform analysis on one channel at a time. For applications such as population screening or DNA sequencing, such a single channel observation and analysis results in an unacceptable delay for screening many members of a population.

### Summary of the Invention

The invention provides a capillary array electrophoresis (CAE) micro-plate. The micro-plate has an array of separation channels connected to an array of sample reservoirs on the plate. The sample reservoirs are organized into one or more sample injectors. A waste reservoir is provided to collect wastes from sample reservoirs in each of the sample injectors. Additionally, a cathode reservoir is multiplexed with one or more separation channels. An anode reservoir which is common to some or all separation channels is also provided on the micro-plate. Moreover, the distance from the anode to each of the cathodes is kept constant by deploying folded channels. The corners on these turns may be right angle turns or more preferably, smooth curves to improve electrophoretic resolution.

In one aspect, the reservoir layout on the substrate separates the sample reservoirs by a predetermined spacing to facilitate the simultaneous loading of multiple samples.

In another aspect, cathode, anode and injection waste reservoirs are combined to reduce the number of holes  $N$  in the substrate to about  $5/4N$  where  $N$  is the number of samples analyzed.

In another aspect, the separation channels are formed from linear segments.

In another aspect, the separation channels are formed from

curvilinear segments, which may include radial segments.

In yet another aspect, the separation channels span from the perimeter of the plate to the central region of the plate. The separation channels may span the plate in a linear or a radial fashion.

In yet another aspect, a CAE micro-plate assembly is formed using a micro-plate, a reservoir array layer, and an electrode array. The assembly simplifies sample handling, electrode introduction and allows an increased volume of buffer to be present in the cathode and anode reservoirs.

Advantages of the invention include the following. The micro-plate of the present invention permits analysis of a large number of samples to be performed at once on a small device. Moreover, the micro-plate allows samples to be easily loaded while minimizing the risk of contamination. Additionally, the micro-plate is easy to electrically address. Further, the micro-plate supports a wide variety of formats that can provide higher resolution separation and detection of samples, faster separation and detection of samples, or separation and detection of more samples.

Other features and advantages will be apparent from the following description and the claims.

#### Brief Description of the Drawings

The accompanying drawings, which are incorporated in and

constitute a part of the specification, schematically illustrate the present invention and, together with the general description given above and the detailed description given below, serve to explain the principles of the invention.

Fig. 1 is a capillary array electrophoresis (CAE) micro-plate layout.

Fig. 2 is a schematic illustration of the sample injector of Figure 1.

Figs. 3A-3D are illustrations of the operation of the sample injector of Figure 2.

Fig. 4A is an exploded perspective view of a CAE micro-plate assembly.

Fig. 4B is a cross-sectional side view of the CAE micro-plate assembly of Fig. 4A.

Fig. 5 is an illustration of a laser excited galvo-scanner in conjunction with a CAE micro-plate.

Figs. 6A and 6B are images of separations of genetic markers for hereditary hemochromatosis.

Fig. 7 is a plot of electropherograms generated from the images of Figs. 6A and 6B.

Fig. 8 is a second CAE micro-plate layout.

Fig. 9 is a third CAE micro-plate layout.

Fig. 10 is a schematic illustration of a sample injector of Figure 9.

Fig. 11 is an enlarged view of a perimeter portion of the CAE micro-plate layout of Figure 9.

Fig. 12 is an enlarged view of a center portion of the CAE micro-plate layout of Figure 9.

#### Description

Referring now to Figure 1, a capillary array electrophoresis (CAE) micro-plate 10 is shown. The micro-plate 10 has an array of capillaries or separation channels 50 etched thereon. In one embodiment of Figure 1, 48 individual separation channels are etched in a 150 micron ( $\mu\text{m}$ ) periodic array. In this embodiment, the separation channels 50 branch out to an 8x12 array of sample reservoirs 101, each of which is spaced a predetermined distance apart to facilitate loading with an 8-tipped pipetter. In this case, each sample reservoir 101 is spaced in one dimension nine millimeters apart from another sample reservoir. The separation channels 50 extend by a first predetermined distance from an injection region to an anode reservoir 180 and by a second predetermined distance from an injector group 100 to a cathode reservoir 120. The first predetermined distance may be about 10 centimeters, while the second predetermined distance may be about 1.8 centimeters.

Each of the sample reservoirs 101 belongs to an injector group such as one of injector groups 100-116. Additionally, injector groups 100, 102 and 104 are connected to a cathode reservoir 120. Although the cathode reservoir 120 is connected to three sample injectors 100, 102 and 104, other cathode injectors may be connected to more than three sample injectors. For instance, a cathode injector 130 is connected to sample

injectors 106, 108, 110, 112, 114 and 116.

The anode reservoir 180 is placed in a non-symmetrical manner in this case to avoid a conflict with a scanning system. Moreover, the distance for paths from the anode reservoir 180 to any one of cathodes 120 or 130 is identical for all separation channels. The equal distance is achieved by providing folded paths connecting certain sample reservoirs that are close to the anode 180 to increase the path length and to achieve a uniform distance between the anode reservoir 180 and the cathode reservoirs 120 and 130 for all sample reservoirs.

In the embodiment of Figure 1, the number of holes H in the micro-plate 10 is about  $5N/4$ , and more exactly,  $5N/4+7$ , where N is a number of samples. As the embodiment of Figure 1 addresses 96 samples in parallel, 127 holes are required to be drilled. This number of holes is close to a theoretical minimum number of holes of  $N+3$ . The reduction in hole counts is advantageous as fewer holes need to be drilled into the micro-plate 10, thereby increasing manufacturing efficiency as well as decreasing the potential for defects in the production of micro-plates, as caused by mechanical stress associated with the drilling process. Another reason for multiplexing the cathode, anode and waste reservoirs is to make it more feasible to fit 96 separation system on a single substrate. The above advantages are also applicable in the event that the holes are formed by a molding process or a bonding process in lieu of the drilling process.

Turning now to Figure 2, details of the sample injector 100

of Figure 1 are shown. The sample injector 100 has a plurality of sample reservoirs 200, 204, 220 and 224. Sample reservoirs 200 and 220 contain a first sample, while sample reservoirs 204 and 224 contain a second sample.

The sample injector 100 also has a first separation channel 202 and a second separation channel 222. The sample injector 100 thus permits a serial analysis of two different samples on each separation channel. The first and second separation channels 202 and 222 are connected to a waste reservoir 208 by a cross channel 207. The sample injector 100 also has a cathode end 210 as well as an anode end 212. The cathode and anode ends 210 and 212 are at opposite ends of the first separation channel 202. Similarly, a second cathode end 214 is connected to a second anode end 216 by a separation channel 222 that is connected to the waste reservoir 208. As illustrated below, by a proper biasing of the anode reservoirs 211 and 212, cathode reservoirs 200 and 214, sample reservoirs 200, 204, 220, 224, and waste reservoir 208, samples may be moved from their respective sample reservoirs 200, 204, 220 and 224 through the cross channel to the waste reservoir thereby facilitating an insertion into the separation channel.

Referring now to Figures 3A, 3B, 3C and 3D, a process for loading a sample from its respective sample reservoir into the cross channel and then performing a separation is shown. In Figure 3A, an injection voltage, preferably about 300 volts (3.0 V/cm), is applied between the sample reservoir 200 and the injection waste reservoir 208 to draw a sample through a channel

that passes from the sample reservoir to the waste reservoir and crosses the separation channel.

In Figure 3B, a separation voltage of about 3700 volts (300 V/cm), for example, is applied between the cathode end 210 and the anode end 212. This causes the electrophoretic separation of the sample. In addition, a back-bias of the potential between the sample reservoir 200 and the injection waste reservoir 208 is applied. Preferably, the back biasing voltage is about 720 volts. The back-biasing operation clears excess samples from the injection cross-channel 213. As illustrated in Figure 3B, a 100 $\mu$ m sample plug is injected and any residual sample is pulled away from the injection region to avoid tailing side-effects.

Figures 3C and 3D represents analogous injections from the second sample reservoir 204. Although the embodiment of Figures 2 and 3A-3D operates on two samples, four samples may be injected onto a single capillary without any significant cross-contamination.

The process of etching patterns into a representative microplate is discussed next. In one microfabricated embodiment, Borofloat glass wafers available from Schott Corporation of Yonkers, NY are pre-etched in 49% HF for 15 sec and cleaned before deposition of an amorphous silicon sacrificial layer of about 1500 $\text{\AA}$  in a plasma enhanced chemical vapor deposition (PECVD) system. The wafers are primed with hexamethyldisilazane, spin coated at 5000 rpm with a photoresist such as a 1818

photoresist available from Shipley Corp. of Marlborough, MA. The photoresist is developed in a 1:1 mixture of Microposit developer concentrate available from Shipley and water. The wafers are then soft-baked at 90° C for 30 minutes. The mask pattern is transferred to the substrate by exposing the photoresist to ultraviolet radiation in a Quintel contact mask aligner. The mask pattern is transferred to the amorphous silicon by a CF<sub>4</sub> plasma etch performed in the PECVD reactor. The wafers are etched in a 49% HF solution for about 3 minutes at an etch rate of 7 μm/min to form a final etch depth of 21 μm and channel width of ~60 μm at the bonded surface. The photoresist is stripped and the remaining amorphous silicon is removed in a CF<sub>4</sub> plasma etch. Holes are drilled into the etched plate using a 1.25 mm diameter diamond-tipped drill bit, available from Crystalite Corporation of Westerville, OH. The etched and drilled plate is thermally bonded to a flat wafer of similar size and type in a programmable vacuum furnace. After bonding, the channel surfaces are coated using a coating protocol.

Turning now to Figures 4A and 4B, an exploded view and a cross-sectional side view of a CAE micro plate are shown. In Figure 4A, a CAE micro-plate 302 with etched separation channels 301 and a plurality of reservoirs 303 formed thereon is provided. A reservoir array layer 304 is mounted above the CAE micro-plate 302 to provide additional reservoir space above the reservoirs formed on the micro-plate 302. The presence of the reservoir

array layer 304 increases the volume of buffers in the cathode and anode reservoirs and simplifies sample handling and electrode introduction. Preferably, the reservoir array layer 304 is a one millimeter thick elastomer sheet which makes a watertight seal when it is in contact with the glass micro-plate 302. The reservoir array layer 304 may be an elastomer such as Sylgard 184, available from Dow Corning of Midland, Michigan.

The reservoir array layer 304 is placed onto the micro-plate 302 before the channels are filled with a separation medium. Preferably, the separation medium is 0.75 percent weight/volume hydroxyethylcellulose (HEC) in a 1X TBE buffer with  $1\mu\text{M}$  ethidium bromide. Additionally, the reservoir array 304 fully isolates the reservoirs from each other. The separation channels are pressure filled with a sieving matrix from the anode reservoir 180 until all channels have been filled. The anode and cathode reservoirs 180 and 120 are then filled with a 10X TBE buffer to reduce ion depletion during electrophoresis. The sample reservoirs are rinsed with deionized water. Samples are then loaded from a micro-titer plate using an 8-tipped pipetter.

After the reservoir array layer 304 is positioned on the micro-plate 302, an electrode array 306 is placed above the reservoir array 304. The electrode array 306 is fabricated by placing an array of conductors such as platinum wires through a printed circuit board. Each conductor is adapted to engage a reservoir on the micro-plate 302. Moreover, the wires are electrically connected with metal strips on the circuit board to

allow individual reservoirs of a common type to be electrically addressed in parallel. The electrode array 306 also reduces the possibility of buffer evaporation. The electrode array 306 in turn is connected to one or more computer controlled power supplies.

As shown in Figure 4B, the reservoir array layer 304, when used in conjunction with the micro-plate 302, enlarges the effective volume of the reservoirs originally formed on the micro-plate 302. Moreover, electrodes from the electrode array 306 are adapted to probe the reservoirs on the micro-plate 302 and the reservoir array layer 306. The solutions are placed in the reservoirs by a pipetter 308.

After assembly, the CAE micro-plate 302 is probed with a galvo-scanner system 400, as shown in Figure 5. The system 400 measures fluorescence using a detector at a detection zone of the channels. During the process of electrophoresis, as a fluorescent species traverses a detection zone, it is excited by an incident laser beam. In a direct fluorescence detection system, either the target species is fluorescent, or it is transformed into a fluorescent species by tagging it with a fluorophore. The passing of the fluorescent species across the detection zone results in a change, typically an increase in fluorescence that is detectable by the system 400.

Turning now to the analysis system, the galvo-scanner 400 has a frequency-doubled YAG laser such as YAG laser available from Uniphase Corporation of San Jose, California. The YAG laser

generates a beam which may be a 30 mW, 532 nm beam. The beam generated by the laser 402 travels through an excitation filter 404 and is redirected by a mirror 406. From the mirror 406, the beam travels through a beam expander 408. After expansion, the beam is directed to a dichroic beam splitter 410. The laser beam is directed to a galvanometer 420 which directs the beam to a final lens assembly 422. In this manner, the beam is focused on a spot of about 5  $\mu\text{m}$  where it excited fluorescence from the molecules in the channels and is scanned across the channels at 40 Hz. The resulting fluorescence is gathered by the final lens and passed through the galvomirror and the dichroic beam splitter 410 to an emission filter 412 which operates in the range of about 545-620 nm. After passing through the emission filter 412, the beam is focused by a lens 414. Next, the beam is directed through a pinhole 416 such as a 400  $\mu\text{m}$  pinhole for delivery to a photomultiplier (PMT) 418.

The electrode array 306 is connected to one or more power supplies 428 such as a series PS300, available from Stanford Research Systems of Sunnyvale, California. The power supplies are connected to a computer and software controlled to automatically time and switch the appropriate voltages into the electrode array 306. The software may be written in a conventional computer language, or may be specified in a data acquisition software such as LabVIEW, available from National Instruments of Austin, Texas. Data corresponding to spatially

distinct fluorescent emission may then be acquired at about 77 kHz using a 16 bit A/D converter from Burr-Brown Corporation of Tucson, Arizona. Logarithmic data compression is then applied to generate five linear orders of dynamic measurement range. The data is obtained as a 16 bit image, and electropherograms are then generated using a suitable software such as IPLab, available from Signal Analytics, Vienna, Virginia, to sum data points across each channel. A detection of all lanes with a 0.09 second temporal resolution has been achieved by the system 400.

## EXPERIMENTS

An electrophoretic separation and fluorescence detection of HFE, a marker gene for hereditary hemochromatosis, was performed to demonstrate the high-throughput analysis of biologically relevant samples using the CAE micro-plates of the present invention. HFE is a genetic disorder that causes a buildup of iron in tissues resulting over time in disease. The buildup primarily affects the liver. Between 0.1 and 0.5% of the Caucasian population are homozygous for an HFE C282Y variant responsible for this disease. If detected early, treatment can be initiated and long term effects avoided. To screen the population for this marker gene, a high throughput screening system is needed.

In this experiment, samples were prepared using PCR

amplification and digestion to assay the C282Y mutation in the HFE gene. This G A mutation at nucleotide 845 creates a *Rsa* I restriction site in the HFE gene. DNA materials were isolated from peripheral blood leukocytes using standard methods. A segment of an HFE exon containing the variant site was amplified with the following primers:

HH-E4B: 5'GACCTCTTCAGTGACCACTC3'

HC282R: 5'CTCAGGCACTCCTCTCAACC3'

The HC282R primer is a primer discussed in Feder et al., Nature Genet. 13, 399-408 (1996), hereby incorporated by reference. The HH-E4B primer contains a 5' biotin tag. The 25  $\mu$ l amplification reaction contained 10 mM Tris-HCl (pH = 8.8), 50 mM KCl, 0.75 mM  $MgCl_2$ , 0.2 mM dNTPs, 7.5 pM of each primer and 1.5 U AmpliTaq DNA, available from Perkin Elmer, Branchburg, NJ. The PCR was carried out under three consecutive conditions: 5 cycles (95° C for 1 min, 64° C for 1 min, 72° C for 1 min), 5 cycles (95° C for 1 min, 60° C for 1 min, 72° C for 1 min), and 25 cycles (95° C for 1 min, 56° C for 1 min, 72° C for 1 min). The restriction digestion of amplified product was carried out by adding 4  $\mu$ l of each amplified sample to 6  $\mu$ l buffer containing 2 U *Rsa* I (Sigma, St. Louis, MO) and digesting for 90 minutes at 37° C. Samples were dialyzed against DI water on a 96 sample dialysis plate, available from Millipore, Bedford, MA. Sample types were initially established by separation of restriction

fragments on 1% Agarose-3% SeaPlaque gel, available from FMC Bioproducts, Rockland, ME, in 0.5x TBE. Gels were stained in 0.5  $\mu$ g/ml ethidium bromide for 30 minutes and visualized on a UV transilluminator, a Spectroline model TR-302, using a 123-bp ladder, available from Life Technologies Inc., Gaithersburg, MD, to determine fragment sizes.

Figure 6A and 6B present images of separations of 96 HFE samples on a CAE micro-plate. The 96 samples were separated in two runs of 48 samples, corresponding to two injection reservoirs per channel. In this experiment, nineteen different samples were dispersed among the 96 sample wells, giving a 5-fold redundancy in sample analysis. An original image 500 was obtained for the first injection, while an original image 504 was obtained for the second injection. Additionally, expanded images 502 and 506, corresponding to original images 500 and 504 are shown. The width of the electrophoretic image shown is 7.4 mm for 48 lanes and the complete analysis of 96 samples was performed in under 8 minutes. The expanded images show that the bands are of high intensity and resolution. The image exhibits a smile with the right lanes about 20 seconds faster than the left. This is caused by a gradient in the electrophoresis voltages caused by the placement of the anode to the side of the injection region to ensure adequate clearance from the scanning lens.

Figure 7 presents the 96 electropherograms obtained from the

images in Figures 6A and 6B. All electropherograms have been shifted to align a 167-bp doublet in order to compare the separations. The 167-bp fragment appears as a doublet due to a partial biotinylation of the HH-E4B primer, as the biotinylated form accounts for the slower migrating fragment in the doublet. The 167-bp doublet provides a useful reference point for the alignment of electropherograms to compare separations and allows an accurate genotyping without requiring a sizing ladder. As shown in Figure 7, an average distance between the 111 and 140-bp bands is 7.3 seconds with a standard deviation (SD) of 0.8 second and 0.6 second, respectively, for the first injection and 6.6 sec with a SD of 1.1 second and 0.5 second, respectively, for the second injection. Using a t-test, the typings for both injections are determined to be at about a 99.9% confidence level.

Referring to Figure 8, a second embodiment of the CAE micro-plate 600 is shown. In Figure 8, the micro-plate 600 is an array of injectors, each of which includes waste reservoirs 602 and 608, sample reservoirs 604, 606, 610 and 612. Each injector unit is connected to one of two cathode reservoirs 614 or 616, respectively. Additionally, each injector unit is connected to one capillary in an array of capillaries or channels 620. The capillaries or channels 620 are connected to an anode 630. In this design, 96 samples can be analyzed by injecting four samples

serially on a single capillary. Further, 24 separation capillaries or channels are used to analyze the material in 96 sample reservoirs. Moreover, each of the injector units has two waste reservoirs. In total, the embodiment of Figure 8 has a hole count of  $3N/2 + 3$ .

Referring now to Figure 9, a third embodiment of the CAE micro-plate 650 is disclosed. In the CAE micro-plate 650 of Figure 9, cathode reservoirs 652 are positioned on a perimeter of the CAE micro-plate 650. Additionally, an anode reservoir 660 is positioned in the center of the CAE micro-plate 650. Separation channels or capillaries may emanate from an outer perimeter of the micro-plate 650 toward the center of the micro-plate 650 in a spiral pattern if longer separation channels are desired. Alternatively, if short paths are desired, the separation channels or capillaries may simply be a straight line connecting the perimeter of the micro-plate 650 to the center 660 of the CAE micro-plate 650.

Turning now to Figures 10 and 11, an injector unit of the CAE micro-plate of Figure 9 and its position on a perimeter of the micro-plate of Figure 9 are illustrated in detail. In Figure 10, two separation channels or capillaries 670 and 671 are connected to a common waste reservoir 672 and a common cathode reservoir 674. Additionally, the separation channels 670 and 671 are connected to sample reservoirs 676 and 678. As shown in

Figures 10 and 11, the connections between the sample and waste reservoirs may intersect in an off-set manner.

Referring now to Figure 12, the common anode 660 of Figure 9 is illustrated in detail. As shown in Figure 11, a plurality of separation channels or capillaries 800-810 form a curvilinear pattern, which may be a radial pattern, converging on a central region 820. From the central region 820, the separation channels or capillaries form a passageway from the perimeter of the central region 820 to the common anode reservoir 660 at the center of the CAE micro-plate. The center area 820 is the area where a rotating scanner may be used for detection purposes.

Samples may be loaded manually or automatically. Serial injections may be used to increase the sample throughput with a predetermined number of capillaries. Moreover, while one embodiment of the present invention injects two samples per channel, an injection of four samples per channel may be used to analyze 192 samples per plate. Further, an increase in the number of capillaries on the CAE micro-plate would increase the throughput correspondingly without introducing any sample contamination. Moreover, the plate may be made of glass or plastic.

In addition, the scanning detection system may be altered by inverting its objective lens and scanning from below. Placing of the optics below the plate would permit facile manipulation and

introduction of samples. The inverted scanning would also avoid spatial conflict with the anode reservoir, thereby permitting a central placement of the anode. Moreover, an array of PCR reaction chambers may be used with the micro-plate of the invention to allow for integrated amplification of low volume samples, eliminate sample handling and manual transfer, and reduce cost. Furthermore, the present invention contemplates that electronic heaters, thermocouples and detection systems may be used with an array of microfluidic capillaries to enhance the CAE electrophoresis process.

While the invention has been shown and described with reference to an embodiment thereof, those skilled in the art will understand that the above and other changes in form and detail may be made without departing from the spirit and scope of the following claims.

What is claimed is:

1           1.    A capillary array electrophoresis plate, comprising:  
2           an array of separation channels formed on said plate; and  
3           an array of sample reservoirs formed on said plate and  
4           coupled to said separation channels.

1           2.    The plate of claim 1, wherein said array of sample  
2           reservoirs are organized into one or more sample injectors.

1           3.    The plate of claim 2, further comprising a waste  
2           reservoir positioned in each sample injector.

1           4.    The plate of claim 3, wherein one of said waste  
2           reservoirs is coupled to one or more sample reservoirs in each  
3           sample injector.

1           5.    The plate of claim 1, further comprising a cathode  
2           reservoir, said cathode reservoir being connected to one or more  
3           separation channels.

1           6.    The plate of claim 1, further comprising an anode  
2           reservoir common to one or more separation channels.

1           7.    The plate of claim 1, wherein the plate has one set of  
2           reservoirs positioned near an outer perimeter, and one set of  
3           reservoirs positioned near a center and the separation channels  
4           connect the reservoirs near the outer perimeter to reservoirs

1 near the center.

1 8. The plate of claim 7, wherein the separation channels  
2 radially connect the outer perimeter to the center.

1 9. The plate of claim 1, further comprising an electrode  
2 array coupleable to said reservoir array.

1 10. The plate of claim 9, further comprising a reservoir  
2 array layer having an array of openings coupleable to said  
3 reservoir array.

1 11. The plate of claim 1, wherein said reservoir array is  
2 regularly spaced in one or two dimensions on said plate and  
3 adapted to engage a multi-headed pipetter.

1 12. A capillary array electrophoresis plate, comprising:  
2 a plurality of separation channels formed at a surface of  
3 said plate;

4 one or more anode reservoirs formed at a surface of said  
5 plate; and

6 one or more injectors formed at a surface of said plate,  
7 said injector having:

8 a plurality of sample reservoirs formed on said plate  
9 and coupled to said separation channels;

10 a plurality of waste reservoirs formed on said plate

1 and coupled to said separation channels; and

2. at least one cathode reservoir multiplexed with a plurality of said separation channels.

1 13. The plate of claim 12, further comprising an electrode array coupleable to said reservoirs.

1 14. The plate of claim 12, wherein the plate has an outer perimeter and a center and the separation channels connect the outer perimeter to the center.

10 15. A capillary array electrophoresis plate comprising:

11 an array of microfabricated separation channels formed at a surface of a first microfabricated substrate and a corresponding surface of a second substrate bonded to said first and second substrates, each of said channels having first and second ends;

13 an array of sample reservoirs formed at a surface of said plate;

14 an array of waste reservoirs formed at a surface of said plate;

15 an array of cathode reservoirs coupled to the first end of each of the separation channels;

16 an array of anode reservoirs coupled to the second end of each of the separation channels; and

17 an injector formed by an injection channel connected to one or more sample reservoirs that crosses a separation channel and

10 connects to a waste reservoir.

1 16. The capillary array electrophoresis plate of claim 15,  
2 wherein both substrates are microfabricated.

1 17. The capillary array electrophoresis plate of claim 15,  
2 wherein the substrates are made of glass.

1 18. The capillary array electrophoresis plate of claim 15,  
2 wherein the substrates are made of plastic.

1 19. The capillary array electrophoresis plate of claim 15,  
2 wherein one or more separation channels are connected to a common  
3 cathode reservoir.

1 20. The capillary array electrophoresis plate of claim 15,  
2 wherein one or more separation channels are connected to a common  
3 waste reservoir.

1 21. The capillary array electrophoresis plate of claim 15,  
2 wherein one or more separation channels are connected to a common  
3 anode reservoir.

1 22. The capillary array electrophoresis plate of claim 15,  
2 wherein one or more sample reservoirs are connected to one  
3 separation channel and one or more waste reservoirs.

1 23. The capillary array electrophoresis plate of claim 15,  
2 further comprising a reservoir array layer mounted above the  
3 plate, the reservoir array layer having openings positioned to  
4 couple to the sample reservoirs, the waste reservoirs, the  
5 cathode reservoirs, and the anode reservoirs.

6  
7 24. The plate of claim 15, further comprising an electrode  
8 array coupleable to said reservoir array layer.

9 25. The capillary array electrophoresis plate of claim 15,  
10 wherein the first substrate has an array of electrodes aligned  
11 with the sample reservoirs, the waste reservoirs, the cathode  
12 reservoirs, and the anode reservoirs to make electrical contacts  
13 with the solutions in the reservoirs.

14 26. The capillary array electrophoresis plate of claim 24  
15 wherein said electrode array is integral with the two substrates.

16 27. The capillary array electrophoresis plate of claim 26,  
17 wherein the sample reservoirs are regularly spaced on the plate  
18 to receive solutions from a multi-headed pipetter system.

19 28. The capillary array electrophoresis plate of claim 15,  
20 wherein the plate has H holes, and wherein H is approximately  
21 equal to  $5N/4$ , with N being the number of samples to be

processed.

29. The capillary array electrophoresis plate of claim 15, wherein the distance from each cathode reservoir to a corresponding injector is approximately equal and where the distance from each injector to its corresponding anode reservoir for each separation channel is approximately equal.

30. The capillary array electrophoresis plate of claim 15, wherein the plate is made of glass or plastic.

31. A method of forming a capillary array electrophoresis plate, comprising:

forming an array of microfabricated separation channels at a surface of the plate;

forming an array of microfabricated sample reservoirs at a surface of the plate; and

connecting the array of microfabricated sample reservoirs to the array of microfabricated separation channels.

32. The method of claim 31, further comprising grouping the array of sample reservoirs into one or more injectors.

33. The method of claim 32, further comprising forming a waste reservoir in each sample injector.

1 34. The method of claim 33, further comprising multiplexing  
2 a cathode reservoir with the sample reservoirs.

1 35. The method of claim 34, further comprising multiplexing  
2 an anode reservoir to all sample reservoirs on the plate, wherein  
3 a distance from each cathode reservoir to a corresponding  
4 injector is approximately equal and where the distance from each  
5 injector to its corresponding anode reservoir for each separation  
6 channel is approximately equal.

1 36. A method for injecting a sample through a capillary  
2 array electrophoresis plate with microfabricated separation  
3 channels connected to sample reservoirs, waste reservoirs,  
4 cathode reservoirs, and anode reservoirs, the method comprising:

5 applying an injection voltage between a first reservoir and  
6 a waste reservoir to draw the sample into a cross channel region  
7 while applying a bias voltage to the cathode and anode reservoirs  
8 to control injection plug width;

9 applying a running voltage between the cathode and anode  
10 reservoirs; and

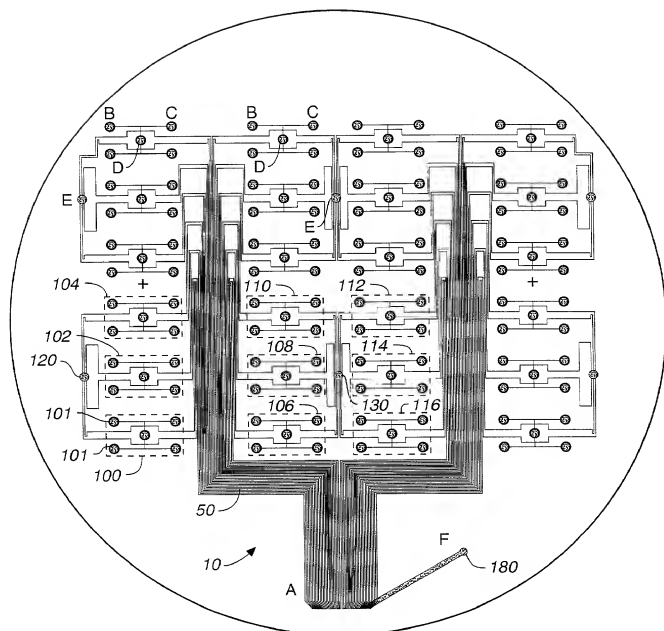
11 applying a biasing voltage to the waste and injector  
12 reservoirs to pull away residuals of the sample.

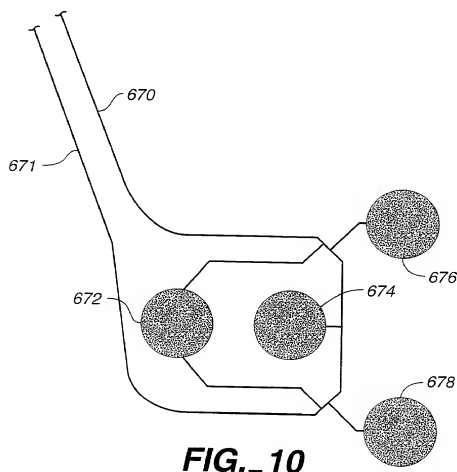
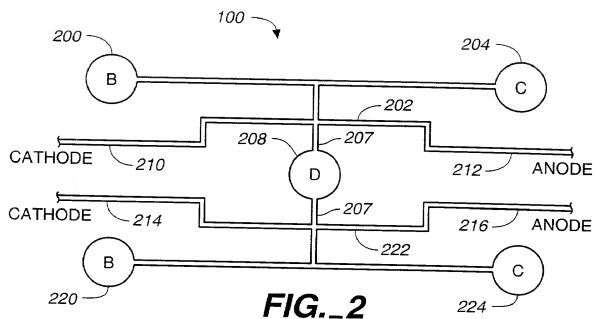
## MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS DEVICE AND METHOD

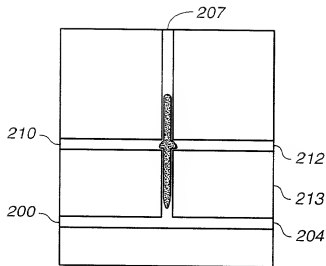
### Abstract

A capillary array electrophoresis (CAE) micro-plate with an array of separation channels connected to an array of sample reservoirs on the plate. The sample reservoirs are organized into one or more sample injectors. One or more waste reservoirs are provided to collect wastes from reservoirs in each of the sample injectors. Additionally, a cathode reservoir is also multiplexed with one or more separation channels. To complete the electrical path, an anode reservoir which is common to some or all separation channels is also provided on the micro-plate. Moreover, the channel layout keeps the distance from the anode to each of the cathodes approximately constant.

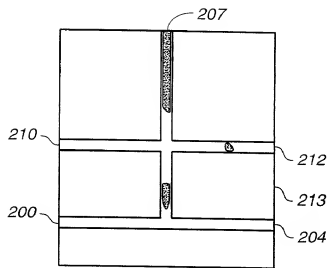
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**FIG. 1**

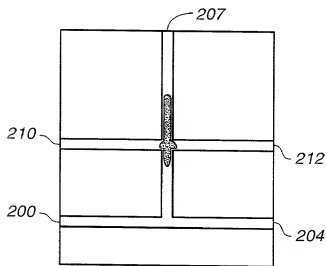




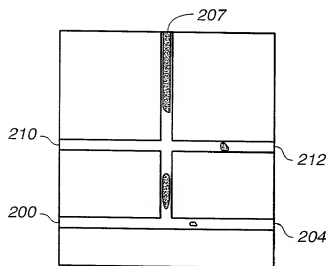
**FIG.\_3A**



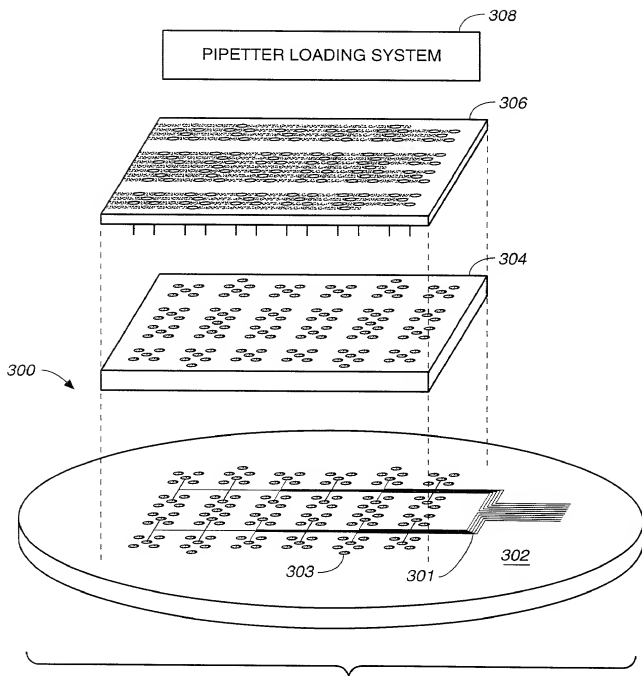
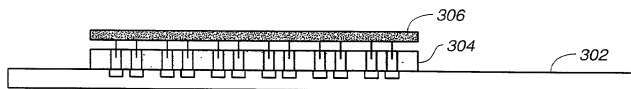
**FIG.\_3B**

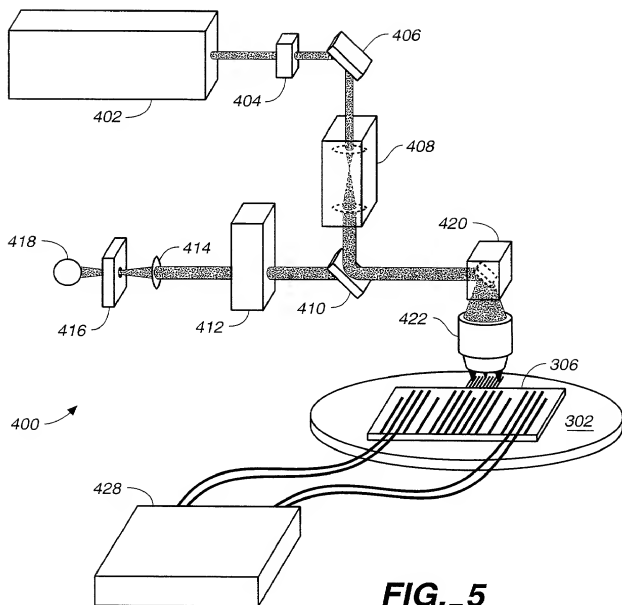


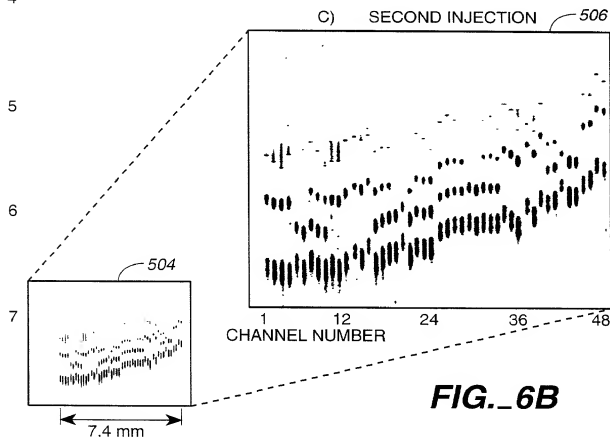
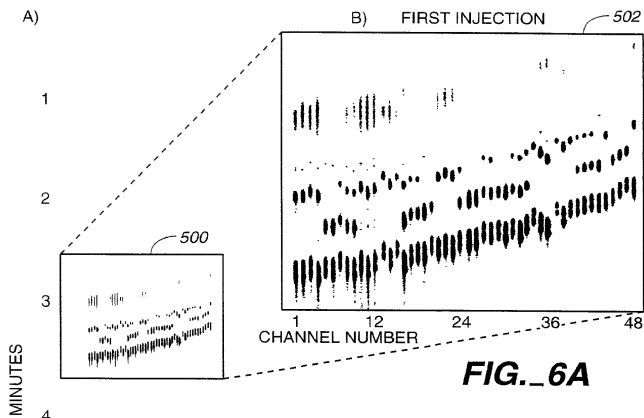
**FIG.\_3C**



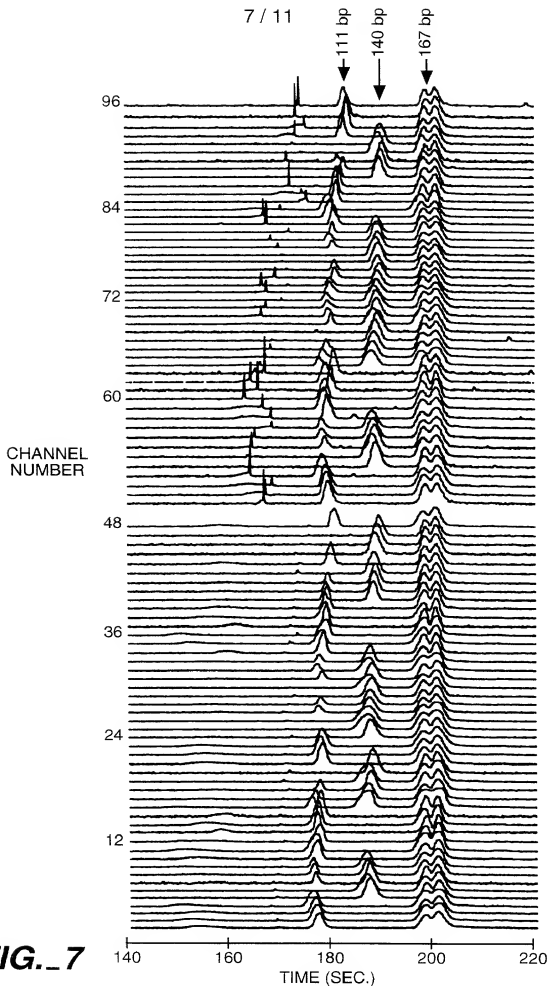
**FIG.\_3D**

**FIG.\_4A****FIG.\_4B**

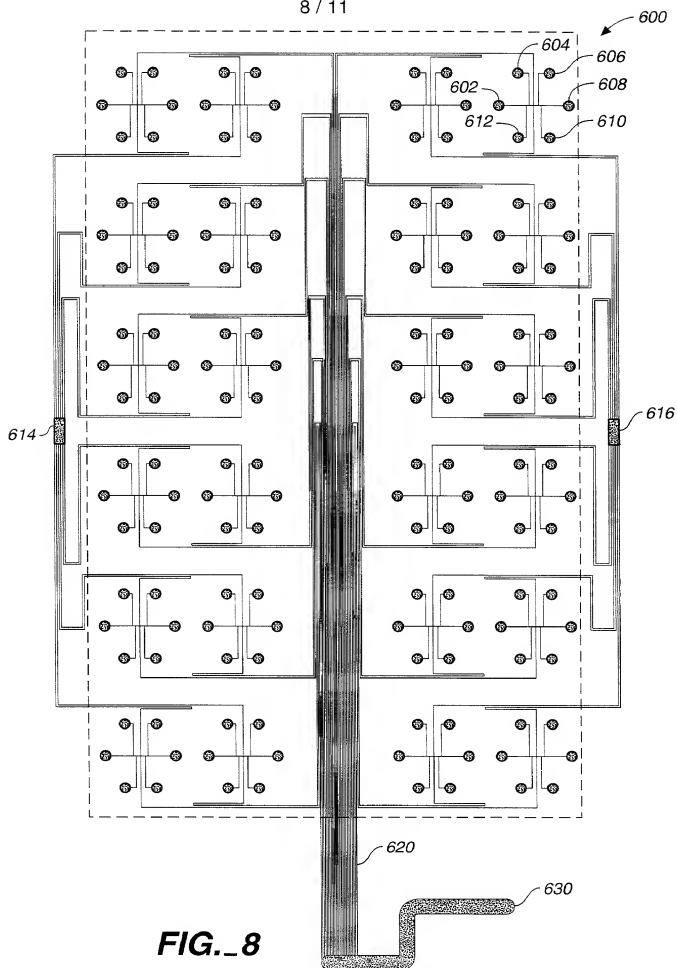


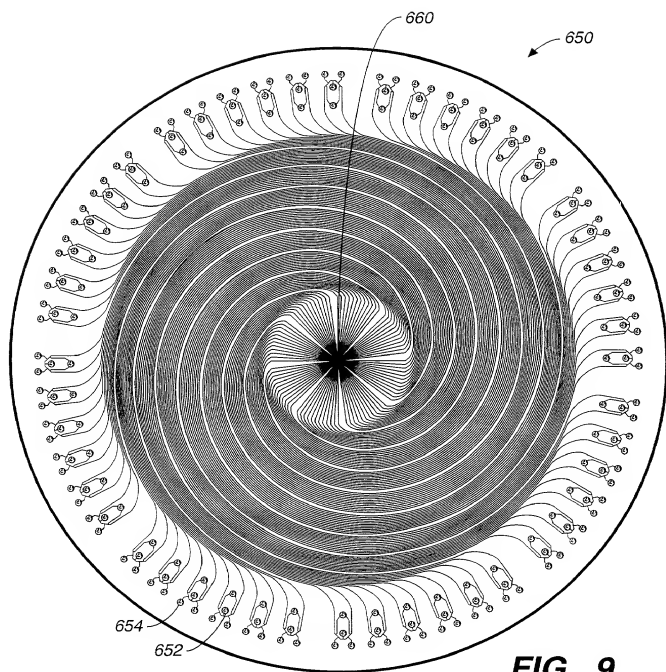


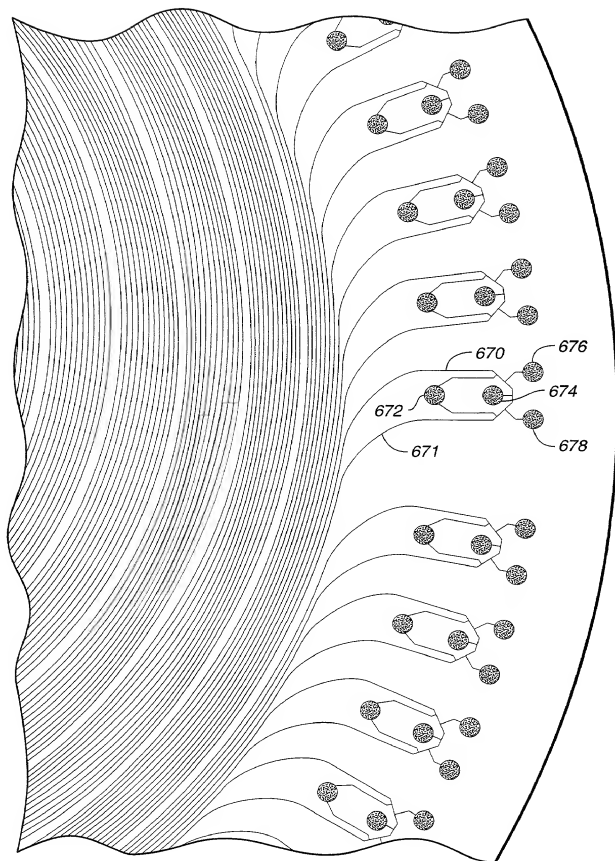
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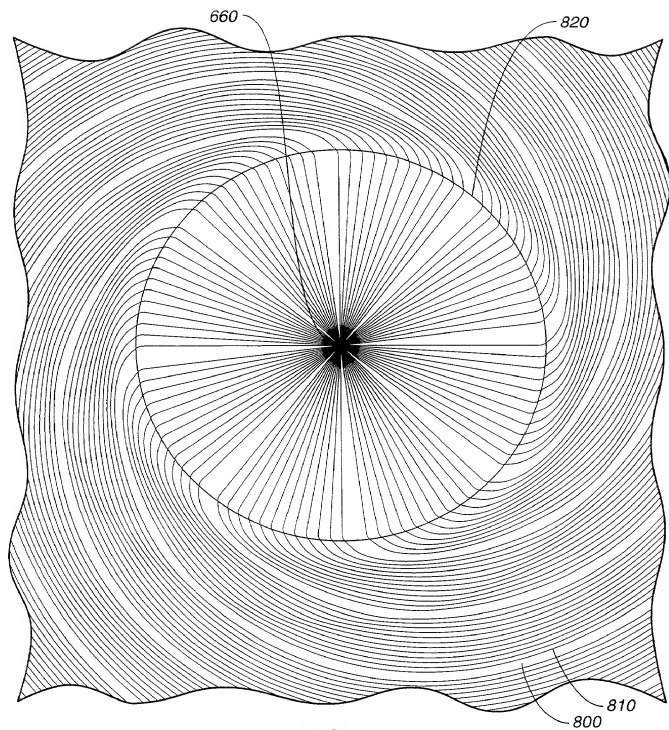


**FIG.\_7**

**FIG.\_8**

**FIG. 9**

**FIG. 11**

**FIG.\_12**

U.C. Case No. B98-020-1

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Peter C. Simpson, et al.  
Serial No.: 08/965,738  
Filed : November 7, 1997  
Title : MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS  
DEVICE AND METHOD

Assistant Commissioner for Patents  
Washington, DC 20231

DECLARATION BY THE INVENTOR(S)

As a below-named inventor(s), I hereby declare that:

My residence, post office address and citizenship are  
as stated next to my name.

I believe I am an original and joint inventor of the  
subject matter which is claimed and for which a patent is sought  
on the invention entitled MICROFABRICATED CAPILLARY ARRAY  
ELECTROPHORESIS DEVICE AND METHOD, the specification of which

☐ is filed concurrently herewith.

☒ was filed on November 7, 1997, as Application  
Serial No. 08/965,738.

I hereby state that I have reviewed and understand the  
contents of the above-identified specification, including the  
claims.

I acknowledge the duty to disclose information which is  
material to the examination of this application in accordance  
with Title 37, Code of Federal Regulations, Section 1.56.

Please direct all correspondence and telephone calls to  
William J. Egan, III, Reg. No. 28,411 Fish & Richardson P.C.,  
2200 Sand Hill Road, Suite 100, Menlo Park, California 94025,  
(650) 322-5070.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first and joint inventor: Peter C. Simpson

Inventor's signature Peter C. Simpson  
Date: April 10, 1998  
Residence: Oakland, California  
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IN THE U.S. PATENT AND TRADEMARK OFFICE

Inventors : Peter C. Simpson, et al.  
Serial No.: 08/965,738  
Filed : November 7, 1997  
Title : MICROFABRICATED CAPILLARY ARRAY ELECTROPHORESIS DEVICE AND METHOD

POWER OF ATTORNEY BY ASSIGNEE AND ELECTION OF  
ASSIGNEE TO CONDUCT PROSECUTION TO EXCLUSION OF INVENTOR(S)

The undersigned, as authorized representative of the assignee of the entire right, title and interest in the above-identified application, hereby appoints


William J. Egan, III, Reg. No. 28,411; David J. Goren, Reg. No. 34,609; Valeta Gregg, Reg. No. 35,127; Hans R. Troesch, Reg. No. 36,950; Wayne P. Sobon, Reg. No. 32,438; Reginald J. Suyat, Reg. No. 28,172; Bao Q. Tran, Reg. No. 37,955

as its attorneys to prosecute the application and to transact all business in the Patent and Trademark Office connected therewith with full powers of substitution and revocation, said appointment to be to the exclusion of the inventor(s) and his attorney(s) in accordance with the provisions of 37 C.F.R. § 3.71 *et seq.* of the Patent Office Rules of Practice.

Ownership is in the assignee by virtue of the assignment documents filed concurrently herewith. The documents evidencing ownership have been reviewed and to the best of the assignee's knowledge and belief, title is in the assignee.

Please direct all communications regarding the application to the attorney at the address and telephone numbers indicated below.

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may 4, 1998   
Date \_\_\_\_\_  
William A. Hoskins  
Director

Assignee: THE REGENTS OF THE UNIVERSITY OF CALIFORNIA  
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